

TITLE: ESPH TRANSLOCATION ASSAY: Name: Rachana Ramachandran

1) Seed HeLa cells in a 6-well plate.

When cells will reach ~70% confluency, infect the cells for 30 min (after 3h preactivation) with the following bacterial strains: 1). $\Delta espH$, 2). $\Delta espH$ +EspH-6xHis-SBP (-IPTG); 3). escV+EspH-6xHis-SBP (+IPTG); 4). $\Delta espH$ +EspH-6xHis-SBP (+IPTG). Induce EspH expression with 0.05 mM IPTG (For EspH, induction with 0.05mM IPTG yielded the highest effector protein expression).

IMPORTANT – Steps 2-9 should be done in the cold/ice.

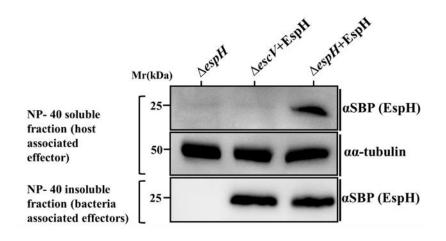
- 2) Wash infected cells 3x with ice-cold 1x PBS. After the last wash, aspirate the buffer as much as possible.
- 3) Add 0.2ml of ice-cold solubilization buffer to each well.
- 4) Scrape of the cells with 'Rubber Policeman' and transfer the lysate to a new ice-cold Eppendorf tube.
- 5) Leave on ice for 1 min
- 6) During that 1 min, mix the lysate by up and down pipetting 5-10 times gently with blue or yellow tip (w/o foaming).
- 7) Spin for $\frac{2 \text{ min}}{2 \text{ min}}$ at 4°C at max g (16,000g)
- 8) Collect the supernatant as much as possible while avoiding the pellet, and place it in an ice-cold Eppendorf tube. It is important to collect all the sup, because it contains the translocated effector, while the pellet contains the bacteria-associated effector.
- 9) Resuspend the pellet by multiple rigorous finger flicking until it is smeared on the wall of the tube.
- 10) Resuspend the pellet and supernatant in 100 μ l of 4x SDS-PAGE sample buffer containing DTT/ β -mercaptoethanol.
- 11) Heat 95 °C for 5 min. For the pellet, sample buffer is added directly to the tube and boiled while for the supernatant, only the required amount to run in a gel is taken (10 or 15μ l) at a time mixed with the sample buffer and boiled. The rest of the Supernatant is aliquoted and stored without adding sample buffer.

ESPH TRANSLOCATION ASSAY:

12) Run 13% gel; Western blot with anti-SBP to detect EspH in the two fractions. Probe the Supernatant (i.e. the NP-40 soluble fraction) with antitubulin for evaluating lysis levels.

Solubilization buffer

- 0.5% NP-40
- 10 mM Tris-HCl (pH-7.4)
- 100 mM NaCl
- 5 mM MgCl₂
- Protease inhibitor cocktail



Ramachandran et al IAI 2018